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INTRODUCTION

During the progression of human PCa hemidesmosomes, adhesion structures that anchor epithelial cells to basement membrane and function as a tumor suppressor, are lost (1, 2). We found that 12lipoxygenase directly interacts with $\beta4$ integrin, an integral part of hemidesmosomes (3, 4). We hypothesized that an increase in 12activity can cause the disassembly of hemidesmosomes, mobilization of $\alpha6\beta4$ integrin from hemidesmosomes to other parts of the cell membrane, and stimulate tumor invasive growth. evaluate proposed to conduct а correlation study to relationship between 12-LOX expression and dispersion of integrin in clinical tumor specimens. Our proposal further aims to study whether 12(S)-HETE, the enzymatic product of 12-LOX, can disrupt hemidesmosomes and whether 12-LOX inhibitors promote the formation of hemidesmosomes. Then we will study the underlying signaling pathway, especially $PKC\alpha$, initiated by 12(S)-HETE, in the disassembly of hemidesmosomes. Next we will overexpress $\beta 4$ integrin and study the role of the interaction between 12-LOX and integrin in the adhesion, proliferation, migration, survival, in response to HGF/SF. Finally we will xenograft these transfected cells into mice, to evaluate whether any phenotypic changes of tumor cells in vitro can be recapitulated in vivo. The work will significantly advance our understanding about complex process of prostate cancer progression as well as the possible role played by dietary fat in the progression of prostate cancer.

BODY OF PROGRESS REPORT

Task 1.

We have attempted several procedures for immunostaining for 12-LOX. In the previous report, the procedure only worked in frozen human prostate tumor tissue. We now worked out the conditions for immunohistochemical analysis of 12-LOX at the protein level in paraffin-embedded human prostate tumor tissues. As shown in the figure 1, 12-LOX immunoreactivity correlated with tumor grade. Neoplastic glands are weakly, moderately or strongly positive for 12-LOX in hyperplatic glands (Figure 1A), atropic glands (Figure 1B), and in tumor Glenson score 4-6 (Figure 1 C,D).

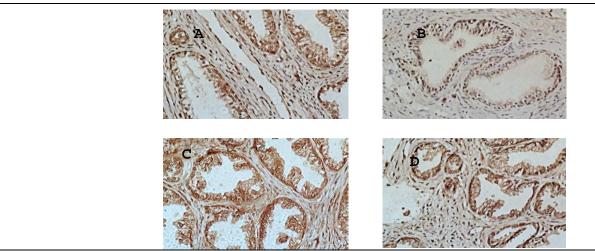


Figure 1. Immunohistochemical staining for 12-LOX in parafirm-embedded prostate tumor tissues. Sections of specimens were probed with a12-LOX polyclonal antibody (Oxford Biomedical Research Inc, Oxford, MI). Positive immunoreactivity is indicated by staining with brownish color. A, hyperplasia gland; B, atropic; C/D, tumor with Glenson score 4-6.

We have also attempted several protocols of immunostaining for $\beta 4$ integrin in paraffin-embedded material. As shown in figure 2, positive staining was found in tumors and correlated with tumor grade.

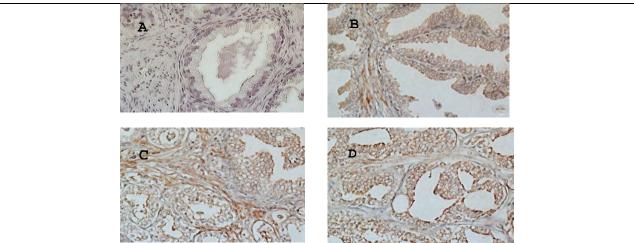


Figure 2. Immunostaining for beta 4 integrin. Brownish color indicates positive staining. Note the pattern of beta 4 integrin positivity. A, normal epithelium; B, tumor with Glenson score 4; C, tumor with Glenson score 6; D, tumor with Glenson score 8.

We have procured 100 cases of prostate tumor specimens. We have analyzed 83 patient samples following the double staining technique. The results are presented in **Table 1**.

Table 1: 12-LOX and b4 Expression in Human Prostate Cancer Compared to Gleason Score

Gleason Score	n	β4 Median (range)	12-LOX Median (range)
6	42	30 (5-80)	15 (5-70)
≥7	41	70 (5-100)	30 (5-80)
p value		0.0082	0.0109

Mann-Whitney test used for statistical analysis. $p \le 0.05$ was considered significant.

Task 2. Study the effects of 12-LOX inhibitors and 12(S)-HETE on hemidesmosome in prostate epithelium. Months 1-18:

We studied the effect of inhibitors of 12-LOX as well as 12(S)-HETE on the hemidesmosome-like structures in cultured cells. As shown in Figures 3 and 4, 12(S)-HETE (300 nM) reduced hemidesmosome like structures both in A431 cells and in an immortalized human prostate cancer cell line, CPTX1532. In contrast, baicalein, a 12-LOX inhibitor, promoted the formation of hemidesmosome-like structure (Figure 4). Interestingly, a linoleic acid metabolite from 15-LOX, 13(S)-HODE, which is known antagonist of 12(S)-HETE, promoted the formation of hemidesmosomes (Figure 3).

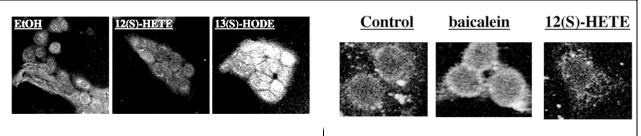
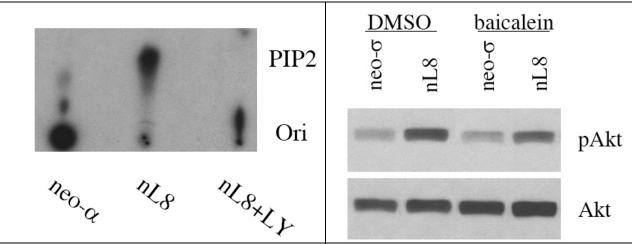


Figure 3: 12(S)-HETE reduced the focal staining of $\beta4$ integrin (middle) while 13(S)-HODE promoted the focal staining (right) in A431 cells. A431 cells were plated on matrigel and treated with 300 nM of 12(S)-HETE or 13(S)-HODE for 30 min, fixed with methanol/acetone, and then processed for immunostaining for $\beta4$ integrin. Note the marked increase in focal staining for $\beta 4$ integrin in 13(S)-HODE treated group (right) vs. the control (EtOH, left). Also note the marked reduction in focal staining for $\beta4$ in 12(S)-HETE treated group (middle) when compared to the control (left).

Figure 4: 12(S)-HETE reduced (right), while 12-LOX inhibitor baicalein promotes (middle), the polarized staining of $\beta4$ integrin in CPTX1532, a human prostate epithelial cell line immortalized with the E6 and E7 transforming proteins of human papilloma virus serotype 16 (Bright et al., 1997). Cells were treated with baicailein or 12(S)-HETE for 30 min, fixed with methanol/acetone, and then processed for immunostaining for $\beta4$ integrin. Note the focal, or polarized staining pattern in baicalein treated group, and less in control group. Cells treated with 12(S)-HETE represented much less focal staining.

Task 3. Study the signal transduction pathways that underlie the disassembly of hemidesmosomes by 12(S)-HETE or an increase in 12-LOX activity, Months 12 -24:



Increased PI3 kinase activity in PC-3 cells by 12-LOX overexpression. PI3 kinase activity in p85 immunoprecipitates was measured by incubating with substrate and γ-32P-ATP. Phospholipids were separated by TLC and autoradiographed. Shown here is a representative of three independent experiments. Note the increased PIP2 production in nL8 when compared to neo and its sensitivity to a PI3 kinase inhibitor, LY294002 (LY). Ori, origin.

Increased Akt phosphorylation in 12-LOX transfected PC-3 cells and its attenuation by baicalein. Neo- α and nL8 cells were treated with DMSO or 15 μ M of baicalein for 16 h. Cell lysates were harvested and the phosphorylation of Akt was analyzed by using a polyclonal antibody against pSer473 Akt (Top panel). Bottom panel, the membrane was stripped and reprobed for total Akt (phosphorylated and nonphosphorylated).

Fig 5. Activation of the PI3K Akt signaling pathway in 12-LOX overexpressing PC-3 cells. A. B.

In Vitro PI3 - Kinase Assay - In vitro phosphorylation of phosphatidylinositols was carried out with PY20 antibody (Transduction laboratory, KY) for the detection of immunoprecipitated PI 3-kinase essentially as previously described (23). Prostate cancer cells were harvested with PBS containing 1 % Nonidet P-40, 1 mM dithiothreitol, 200 μ M sodium vanadate, 100 μ M 4-(2- aminoethyl) benzenesulfonyl fluoride, 10 μ g/ml leupeptin, 10 μ g/ml aprotinin (lysis buffer). Soluble cell lysates were collected after microcentrifugation at 18,000 g for

5 min. The protein concentration of cell lysates was adjusted to 0.8-1 µg/µl with lysis buffer and PY20 mAb (5 µg) was added to 500 µl of the cell lysate. The mixture was incubated with gentle rocking at 4 °C overnight and then 10 mg of protein A -Sepharose (Amersham Pharmacia Biotech) were added and the incubation was continued for another 2 h. The immunoprecipitates were washed (3 x) with the aforementioned lysis buffer, twice with 0.1 M Tris/HCl, pH 7.5, containing 0.5 M LiCl, and 10 μ M sodium vanadate, and twice with 10 mM Tris/HCl, pH 7.5, containing 100 mM NaCl, 10 μ M sodium vanadate, and 1 mM EDTA. Then 50 μ l kinase buffer [10 mM HEPES (pH7.0), 0.1 mM EGTA, 25 mM MgCl2, 100 μ M ATP, 10 μ Ci of [32P]ATP, 10 μ g/sample PtdIns] was added to each sample, and incubated at room temperature for 10 min. The assay was terminated by adding 60 μ l of 2 μ M HCl and 160 µl of chloroform/methanol (1:1) / sample, and followed by vortexing and centrifugation. The chloroform phase which contained the inositides was removed and applied to Silica gel 60 thin layer chromatography (TLC) sheets (Merck). TLC was developed with chloroform : methanol : 24% ammonium hydroxide : H2O (90 : 90 : 9 : 19). After drying, spots were located by autoradiography and compared with standards.

Task 4. Overexpress b4 integrin in PC-3 cells, in the presence or absence of 12-LOX expression, and evaluate the capacity of transfected cells to form hemidesmosomes and whether an increase in surface expression of a6b4 alters cell proliferation, adhesion, migration, and survival, in response to HGF/SF, Months 18 - 30:

Generation of PC-3 cells expressing various $\beta4$ mutants: We constructed a panel of bacterial expression plasmids and mammalian expression constructs that express various b4 mutants of the cytoplasmic tail as shown in Figure 6. Using pGR30~47, we have generated a panel of stable transfectants that express various b4 mutants as GFP fusion proteins through fluorescence activated cell sorting (FACS) and G418 selection for the transfectants.

The expression of $\beta4$ mutants as GFP-fusion proteins is shown in figure 7. Co-immunoprecipitation was performed to determine the interaction of 12-LOX with various $\beta4$ mutants. As shown in the figure 8, 12-LOX is able to bind to the fusion proteins expressed by pGR40, pGR45, pGR46, and pGR47, but not to the fusion protein with only the linker region (pGR38). 12-LOX weakly binds to the fusion protein encoded by pGR39. Collectively, the data suggest a strong binding site(s) for 12-LOX located between 1126-1315 of $\beta4$ cytoplasmic tail.

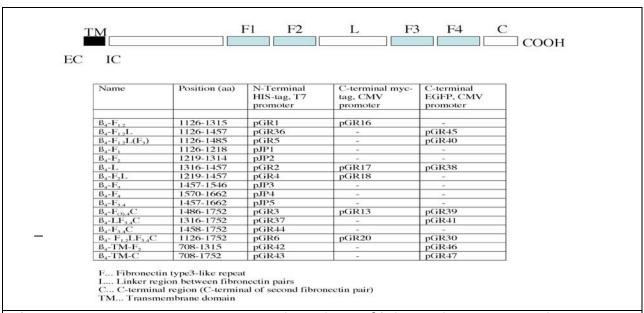
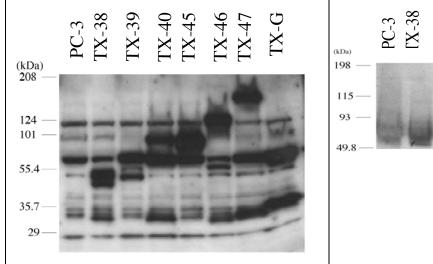


Figure 6. Schemes of the cytoplasmic tail of $\beta4$ integrin and bacterial and mammalian expression constructs. TM, transmembrane; EC, extracellular; IC, intracellular



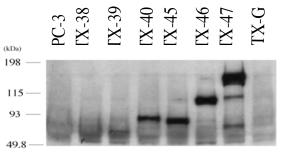
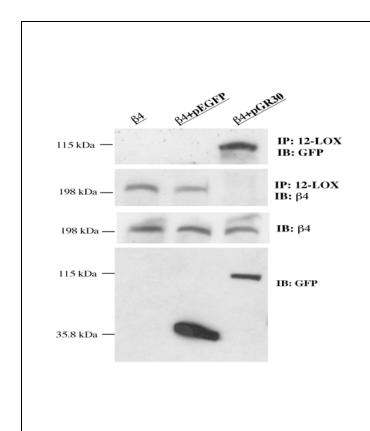


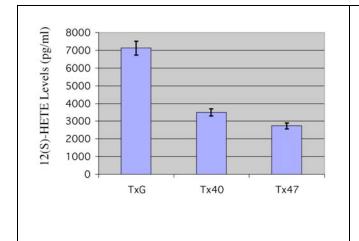
Figure 7. Western blot analysis for the expression of $\beta4$ mutants in PC-3 transfectants with an antibody against GFP. Note the expression of GFP fusion proteins of various MWs. The calculated MW for fusion proteins for pGR47 (Tx47) is about 146. For pGR46 (Tx46), 97.4; pGR45 (Tx45), 66.8; pGR40 (Tx40), 70; pGR39 (Tx39), 59.6; and pGR38 (Tx38), 45.7. The MW for pEGFP (TxG) is approximately 30.

Figure 8. Co-immunoprecipitation of 12-LOX with the fusion proteins expressed by pGR40, pGR45, pGR46, and pGR47, but not to pGR38 and pEGFP (TxG). It is deduced that a binding site or sites are located between 1126 - 1315 region of the $\beta4$ cytoplasmic tail.

Cytosolic $\beta 4$ cytoplasmic tail binds to 12-LOX and blocks the interaction between 12-LOX and full length $\beta 4$ integrin: As shown above, we have constructed a panel of expression constructs encoding various mutants for the $\beta4$ cytoplasmic tail. The construct pGR30 encodes the cytosolic $\beta4$ cytoplasmic tail, with TM domain deleted, as a GFP-fusion protein (Figure 6). When ectopically expressed in 12-LOX transfected PC-3 cells, the cytosolic $\beta4$ cytoplasmic tail interacts with 12-LOX (Figure 9, the IP: 12-LOX / IB: GFP panel). Interestingly the ectopically expressed cytosolic $\beta4$ tail blocks the interaction of 12-LOX with full-length $\beta4$ integrin (Figure 9, the IP: 12-LOX / IB: $\beta4$ panel). The blockade of the interaction between 12-LOX and full-length $\beta4$ integrin is not due to the lack of $\beta4$ expression (Figure 9, the IB: $\beta4$ panel), but due to the presence of the cytosolic β 4 cytoplasmic tail (Figure 9, the IB: GFP panel). The results suggest that the cytosolic $\beta4$ mutant, encoded by pGR30, is able to block the interaction between 12-LOX and full-length $\beta4$ integrin in a dominant negative manner.



Cytosolic Figure 9: β4 encoded by pGR30, binds to 12-LOX blocks the interaction between 12-LOX and full length $\beta4$ integrin. 12-LOX transfected PC-3 cells (PC3/12LOX) transiently transfected with an expression construct for fulllength $\beta 4$, in the presence or absence of pGR30 or its vector, Cell lysates immunoprecipitated with а polyclonal antibody against 12-LOX and immunoblotted for GFP (JL8 mAb, Clontech) or fulllength β4 integrin (AB1922, Chemicon International. band, around 205 kDa). Total lysates were also analyzed for the expression of full-length $\beta4$ integrin and GFP or GFP-fusion protein by immunoblot. Note the co-immunoprecipitation of 12-LOX and the cytosolic $\beta4$ cytoplasmic tail and the blockade of interaction between 12-LOX full-length $\beta4$ integrin by cotransfection with pGR30.



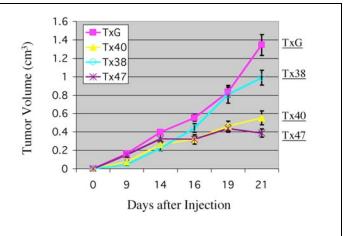


Figure 10. Decrease in 12(S)-HETE biosynthesis in PC-3 cells that express 12-LOX binding $\beta4$ mutants. 12(S)-HETE was isolated from cell lystaes and measured by EIA. N = 6 for each type of cells. Bar, SEM.

Figigure 11. Alteration in PC-3 tumor growth as a result of ectopic expression of 12-LOX binding $\beta4$ mutants (Tx40 and Tx47), not non-12LOX-bind mutant (Tx38). TxG is the vector control. Each datum point, the average tumor volume from 4 mice. Bar, SEM. Note the correlation of the reduction in tumor growth with reduced 12(S)-HETE biosynthesis shown in figure 7.

Task 5. Evaluate the growth rates of s.c. tumors derived from $\alpha6\beta4$ expressing PC-3 cells, in the presence or absence of stable 12-LOX expression, and compare with that of control PC-3 cells, Months 24-36:

We have conducted a preliminary study to determine whether ectopic expression of $\beta4$ mutants, especially those that can bind 12-LOX, can effect 12-LOX activity. As shown in Figure 10, there reduction in 12(S)-HETE biosynthesis in PC-3 cells expressing two 12-LOX binding $\beta4$ mutants (pGR40 and pGR47). We are still in the process of determining whether 12-LOX cellular localization and interaction with the full-length $\beta4$ integrin are altered as result of the presence of $\beta4$ mutants encoded by pGR40 and pGR47. When injected into athymic nu/nu mice, it was found, as shown in Figure 11, that there were a reduction in tumor growth rate in PC-3 cells expressing 12-LOX binding β4 mutants (Tx40 and Tx47, see figure 11), when compared to those from vector control (TxG) or a $\beta4$ fragment that does not interact with 12-LOX (Tx38, see Figure 11).

Key Research Accomplishments:

• Optimized the conditions for immunohistochemical analysis of 12-LOX and $\beta4$ at the protein level in paraffin-embedded human prostate tumor tissues. This standardized method had enabled

- us to continue with the correlation between 12-LOX expression and distribution of $\beta 4$ integrin with Gleason score.
- Constructed several $\beta4$ cytoplasmic tail mutants and identified the sequence of amino acids on $\beta4$ integrin that interacts with 12-LOX.
- Preliminary experiments conducted to show the feasibility of disruption of the interaction of 12-LOX with $\beta4$ integrin in PC-3 cells with ectopic expression of the cytoplasmic tail sequence of $\beta4$ integrin. This resulted in the reduction of the biosynthesis of 12-HETE as well as tumor growth from PC-3 cells in experimental animals.

Reportable Outcomes:

NONE

Conclusions:

Interaction of 12-LOX with the cytoplasmic tail of $\beta4$ integrin results in the disruption of hemidesmosomes. Our data presented here demonstrates the potential to exploit the interaction between 12-LOX and $\beta4$ integrin using ectopically expressed cytoplasmic tail sequence of the integrin to modulate tumor growth.

References:

- 1. Nievers MG, Schaapveld RQ, Sonnenberg A. Biology and function of hemidesmosomes. Matrix Biol 1999;18(1):5-17.
- 2. Jones JC, Hopkinson SB, Goldfinger LE. Structure and assembly of hemidesmosomes. Bioessays 1998;20(6):488-94.
- 3. Tang K, Finley RL, Nie D, Honn KV. Identification of 12-lipoxygenase interaction with cellular proteins by yeast two-hybrid screening. Biochemistry 2000;39(12):3185-91.
- 4. Tang K, Nie D, Cai Y, Honn KV. The $\beta4$ integrin subunit rescues A431 cells from apoptosis through a PI3K/Akt kinase signaling pathway. BiochemBiophysResCommun 1999;264(1):127-32.

Appendices:

NONE